

## INVITED COMMENTARY

**Intracellular lactate flux: a new regulator of the allogenic immune response\***

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\*Invited commentary on "Immunosuppressive properties of a series of novel inhibitors of the monocarboxylate transporter MCT-1", by Pählman *et al.* [*Transpl Int* 2013; 26: 22].

Received: 13 November 2012

Accepted: 15 November 2012

doi:10.1111/tri.12035

During the past decade, long-term graft- and patient-survival rates have not improved significantly, despite the reduced incidence of acute rejections. This is partly because of the toxicities of immunosuppressive drugs and patients' deaths from cardiovascular events, which can be associated with the cardiovascular risk factors related to immunosuppressive drugs [1]. Among these drugs, calcineurin inhibitors (CNIs) are strongly associated with renal toxicity and the development of cardiovascular risk factors, including hypertension, diabetes, and dyslipidemia; thus, there is a need for a toxicity-free CNI-free regimen [2]. However, very few molecules have emerged that can totally or partially replace CNIs. Most evaluated molecules have targeted or regulate T-cell activation, especially signal 1 or 2 activation of T cells, as well as the third signal involved in the expansion of activated T cells.

It has recently been shown that other mechanisms are implicated in the development or differentiation of T lymphocytes. These modify nutrients or precursors that are not only involved in the expansion phase of activated lymphocytes, such as inositol monophosphate dehydrogenase, but are also able to modulate T-cell activation or to switch their phenotype from activated T cells to regulatory cells. For example, tryptophan metabolism induced by IDO-expressing dendritic cells is associated with inducing regulator T cells [3]. Adenosine metabolism, induced by the CD73

ecto-enzyme, also participates in the development of regulatory cells [4]. However, the mechanisms that make these metabolites effective are still not completely understood, although they do affect the phenotype of T cells, which leads to the development of regulatory T cells. For adenosine, the binding of the small molecule induces some signaling in the cell through the A2A adenosine receptor [5]. Their role in organ transplantation has yet to be demonstrated, but this finding opens up a new area of transplantation research.

Other compounds may also be of interest. Pählman *et al.* [6] demonstrated that inhibition of lactate transport in lymphocytes favored tolerance in rats. The lactate transporters are a group of proton-linked monocarboxylate transporters of at least eight members but only four have been functionally characterized (MCT-1, MCT-2, MCT-3, MCT-4) that have been cloned independently by two research groups [7]. MCT-1 is widely expressed *in vivo* in several tissues such as kidney, gut, hematopoietic cells, brain, gut, muscles, and lymphocytes [8,9]. MCT-1 can favor the release of lactate accumulated in cells following glycolysis. Without MCT-1, lactates and pyruvates will be accumulated in the cytoplasm of cells and the normal aerobic glycolysis can't be processed because of the intracellular accumulation of lactate and protons. Overexpression of MCT-1 in Chinese hamster ovary cells or in chronically stimulated skeletal muscles increases

lactate efflux and maintain both glycolysis and function of cells indicating that MCT-1 is a critical lactate-transport protein across membranes [10,11]. Activity patterns may also be important for establishing the capacity of lactate transport in muscle. When muscle activity is reduced by hind-limb unweighting or denervation, lactate transport is decreased. On the other hand, after chronic muscle stimulation and exercise training, lactate transport is increased in endothelial cells. MCT-1 promotes angiogenesis by increasing the lactate activation of HIF1 $\alpha$ , which is implicated in the secretion of proangiogenic factors (vascular endothelial growth factor and fibroblast growth factor) and also increases cell survival by activating the NF $\kappa$ B pathway [12]. MCT-1, but not MCT-2, MCT-3, or MCT-4, is significantly expressed in activated lymphoid cells but not in resting cells probably because activation of T cells required an important production of energy for their multiplication and activation. MCT-1 expression at the plasma membrane is stabilized by CD147 [13]. Interestingly, Murray *et al.* have identified several compounds that inhibit MCT-1 with a high affinity, but not other members of this family [14]. These compounds are able to inhibit lymphocyte proliferation which correlates with lactate accumulation but do not induce cell death. They demonstrated that two inhibitors are efficient at preventing graft rejection. However, they are also associated with side effects, such as testis toxicity, probably because of the expression of MCT-related molecules or MCT-1 in testis cells. By introducing several modifications to the core of the pyrrolopyrimidine of their initial compounds, researchers have now selected molecules that have a similar ability to impair lymphocytic activation and induce graft tolerance in rat heart transplant models in both combinations of low or high responders. The effect was maintained after the withdrawal of the molecule suggesting that some of this compound induce tolerance. This was tested in a situation of second transplantation without any immunosuppressive treatment. The mechanism by which these compounds may induce tolerance in rodents is still unknown.

Surprisingly, these molecules have a slightly lower affinity to MCT-1 but are less lipophilic and have a better bioavailability than the initial compound. After a short period of administration in a small-animal model, a good safety profile has been observed, though extended analyses are needed to demonstrate the absence of other side effects in big animals and humans. In addition, since MCT1 is expressed in the brush border of tubular epithelial cells, and because MCT1 is overexpressed in hypoxic conditions, particular attention has to be done in other transplantation such kidney transplantation in which ischemia-reperfusion may stimulate the expression of MCT-1 and for which inhibition of MCT-1 may lead to important lesion because these cells need a lot of energy for their different functions.

## References

- Callaghan CJ, Bradley JA. Current status of renal transplantation. *Methods Mol Biol* 2006; **333**: 1.
- Casey MJ, Meier-Kriesche HU. Calcineurin inhibitors in kidney transplantation: friend or foe? *Curr Opin Nephrol Hypertens* 2011; **20**: 610.
- Terness P, Bauer TM, Rose L, *et al.* Inhibition of allogeneic T cell proliferation by indoleamine 2,3-dioxygenase-expressing dendritic cells: mediation of suppression by tryptophan metabolites. *J Exp Med* 2002; **196**: 447.
- Romio M, Reinbeck B, Bongardt S, Hüls S, Burghoff S, Schrader J. Extracellular purine metabolism and signaling of CD73-derived adenosine in murine Treg and Teff cells. *Am J Physiol Cell Physiol* 2011; **301**: C530.
- Ohta A, Kini R, Ohta A, Subramanian M, Madasu M, Sitkovsky M. The development and immunosuppressive functions of CD4(+) CD25(+) FoxP3(+) regulatory T cells are under influence of the adenosine-A2A adenosine receptor pathway. *Front Immunol* 2012; **3**: 190.
- Pählman C, Qi Z, Murray CM, *et al.* Immunosuppressive properties of a series of novel inhibitors of the monocarboxylate transporter MCT-1. *Transplant International* 2013; **26**: 22.
- Halestrap AP, Price NT. The proton-linked monocarboxylate transporter (MCT) family: structure, function and regulation. *Biochem J* 1999; **343**: 281.
- Bonen A, Heynen M, Hatta H. Distribution of monocarboxylate transporters MCT1-MCT8 in rat tissues and human skeletal muscle. *Appl Physiol Nutr Metab* 2006; **31**: 31.
- Becker HM, Mohebbi N, Perna A, Ganapathy V, Capasso G, Wagner CA. Localization of members of MCT monocarboxylate transporter family Slc16 in the kidney and regulation during metabolic acidosis. *Am J Physiol Renal Physiol* 2010; **299**: F141.
- Tamai I, Takanaga H, Maeda H, Sai Y, Ogihara T, Higashida H, Tsuji A. Participation of a proton-cotransporter, MCT1, in the intestinal transport of monocarboxylic acids. *Biochem Biophys Res Commun* 1995; **214**: 482.
- McCullagh KJ, Poole RC, Halestrap AP, Tipton KF, O'Brien M, Bonen A. Chronic electrical stimulation increases MCT1 and lactate uptake in red and white skeletal muscle. *Am J Physiol* 1997; **273**: E239.
- Sonveaux P, Copetti T, De Saedeleer CJ, *et al.* Targeting the lactate transporter MCT1 in endothelial cells inhibits lactate-induced HIF-1 activation and tumor angiogenesis. *PLoS ONE* 2012; **7**: e33418.
- Kirk P, Wilson MC, Heddle C, Brown MH, Barclay AN, Halestrap AP. CD147 is tightly associated with lactate transporters MCT1 and MCT4 and facilitates their cell surface expression. *EMBO J* 2000; **19**: 3896.
- Murray CM, Hutchinson R, Bantick JR, *et al.* Monocarboxylate transporter MCT1 is a target for immunosuppression. *Nat Chem Biol* 2005; **1**: 371.