

Protective effects of the lazaroid U74500A and lidoflazine on liver preservation with UW solution

J. Jacobsson, R. Sundberg, H. L. R. Rilo, A. Gasbarrini, T. E. Starzl, D. Van Thiel

¹ Department of Surgery, University of Pittsburgh, 3601 Fifth Avenue, Pittsburgh, PA 15213, USA

Received: 23 September 1992/Received after revision: 5 January 1993/Accepted: 14 January 1993

Abstract. The effect of adding a 21-aminosteroid, U74500A, and a Ca^{2+} antagonist, lidoflazine, alone and together to UW solution was assessed in a rat liver preservation model. Following preservation, the livers were reperfused using a closed circuit, and the release of hepatocellular enzymes (ASAT, ALAT, and LDH) into the perfusate was determined with increasing time. Both drugs reduced the amount of enzymes lost from the liver. The combination of the two drugs was better than either drug alone. These data suggest that both agents may be of value in organ preservation for clinical liver transplantation.

Key words: Liver preservation, rat, UW solution – Rat, liver preservation, UW solution – U74500A, rat liver preservation – Lidoflazine, rat liver preservation

Introduction

Lazaroids are a new group of 21-aminosteroid compounds that have recently attracted interest because of their membrane-stabilizing properties [5, 10, 12]. In particular, they have been shown to reduce iron-dependent lipid peroxidation, which is an important mechanism for oxygen free, radical induced hepatic injury occurring as a consequence of ischemia and reperfusion. These data suggest that these agents may be of value in organ preservation. In the present study, the effect of adding the lazaroid U74500A to University of Wisconsin (UW) solution on the hepatic injury experienced as a result of organ preservation was evaluated using the isolated, perfused liver. The magnitude of the improvement achieved with U74500A was compared to that achieved with a Ca^{2+} antagonist, lidoflazine, which is known to reduce the hepatic injury incurred as a consequence of hepatic preservation using the same model [13]. In addition, the effect of adding both of these drugs to the UW solution was determined.

Correspondence to: R. Sundberg, Karolinska Institute, Department of Transplant Surgery, Huddinge Hospital, S-141 86 Huddinge, Sweden

Materials and methods

Male Lewis rats (Harlan, Indianapolis, Ind.) weighing 240–340 g were used for the experiments. U74500A (Upjohn, Kalamazoo, Mich.), 21.6 mg/l, or lidoflazine (Kabi Pharmacia, La Jolla, Calif.), 5 mg/l, or a combination of both were dissolved in UW solution (DuPont Critical Care, Wilmington, Del.). No insulin, methylprednisolone, or antibiotics were added. The hepatectomy and rat-isolated perfusion technique used were identical to those previously described [13]. Thirty milliliters of the preservation medium was used to flush the portal vein, and the livers were stored in 100 ml of the same solution. Control livers were flushed and stored with UW solution without additives. The livers were stored at 0°C for 72 h before reperfusion was started using a closed circuit, with Krebs-Henseleit bicarbonate solution containing 2% albumin and 5 mM glucose as the perfusate. At 30 and 60 min after reperfusion, samples of the perfusate were taken for analysis of aspartate aminotransferase (ASAT), alanine aminotransferase (ALAT), and lactate dehydrogenase (LDH). The amount of hepatocellular enzymes present in the perfusate was determined using a Technicon RA-500 analyzer.

Results were calculated as means \pm standard deviations (SD). Statistical comparisons were performed using the Wilcoxon rank-sum test. A *p* value less than 0.05 was considered statistically significant.

Results

All livers lost weight during cold storage with no detectable difference between groups [12.8% \pm 2.0% (lidoflazine + lazaroid) vs 10.7% \pm 3.7% (lidoflazine) vs 11.3% \pm 4.9% (lazaroid)]. The amount of ASAT, ALAT, and LDH released into the perfusate during the perfusion period is shown graphically in Figs. 1–3. The levels of all three enzymes increased with increasing time of the reperfusion. The addition of U74500A to UW solution was associated with a significant reduction in the release of ASAT and LDH at 30 and 60 min. The reduction in ALAT release was not significant. The addition of both agents to the UW solution resulted in a significantly reduced initial release of ASAT, ALAT, and LDH into the perfusate medium compared to what was seen with either drug alone. After 60 min of reperfusion, however, the levels of all three enzymes in the perfusate had increased to the level achieved with the addition of either agent alone.

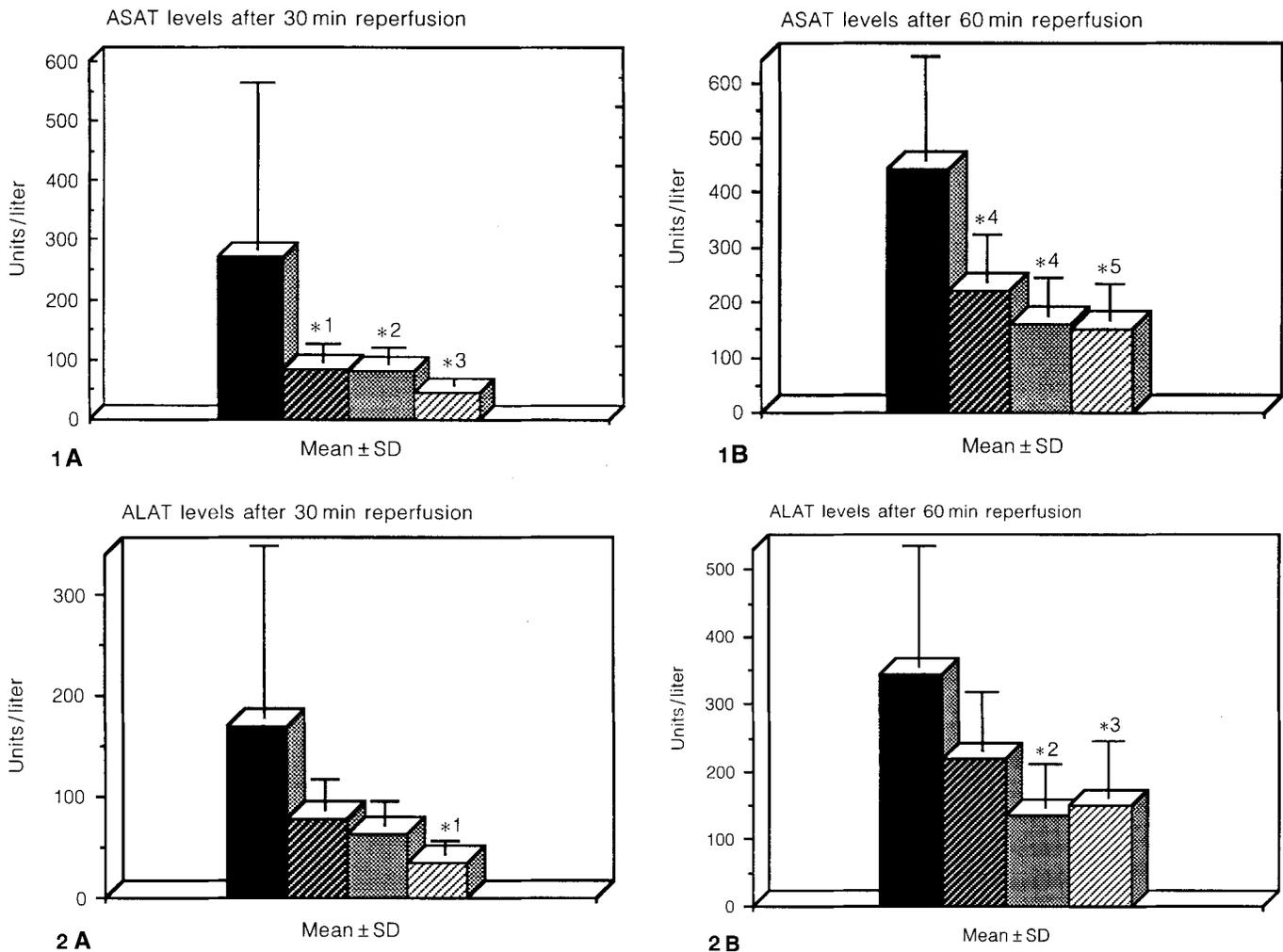


Fig. 1 A, B. The amount of ASAT released into the perfusate was significantly reduced after **A** 30 and **B** 60 min, when U74500A, lidoflazine, or both drugs were added to the preservation medium. At 30 min the drug combination was more effective than either drug alone ($P = 0.016$ for both comparisons), whereas there was no significant difference at 60 min. ■ UW solution, ▨ lazaroïd, ▩ lidoflazine, ▪ lazaroïd + lidoflazine. *1 $P = 0.01$; *2 $P = 0.011$; *3 $P = 0.001$; *4 $P = 0.025$; *5 $P = 0.007$

Fig. 2 A, B. The amount of ALAT released into the perfusate was not significantly altered **A** after 30 min, when U74500A or lidoflazine was added to the preservation medium. **B** After 60 min a reduction was seen in the lidoflazine group, but not with U74500A. The drug combination was found to reduce the release of enzyme at 30 and 60 min. At 30 min the combination was more effective than lidoflazine alone ($P = 0.031$). ■ UW solution, ▨ lazaroïd, ▩ lidoflazine, ▪ lazaroïd + lidoflazine. *1 $P = 0.004$; *2 $P = 0.01$; *3 $P = 0.016$

Discussion

A major principle in organ preservation is the use of hypothermia. Hypothermia reduces the rate of cellular metabolism and thereby the number of various metabolic events that occur during ischemia that lead to cell injury and death. Hypothermia is not without side effects, however, that include cell swelling. This effect can be counter-

acted with the use of a flush solution that contain cell impermeants, such as those present in the UW solution [3, 19]. Organ preservation can be improved further with the use of pharmacological agents that interfere with key processes in the pathogenesis of cell injury occurring as a result of ischemia and reperfusion. Examples of such agents are membrane stabilizers including chlorpromazine [17, 18], glucocorticoids [17], oxygen-free radical scavengers [15], vasodilators [11], and calcium antagonists [2, 13].

Glucocorticoids have been used extensively in experimental studies to reduce injury experienced with trauma, especially neurotrauma [8], ischemia [16] and, in some studies, the injury associated with organ preservation [7, 17]. The putative mechanism behind the protective effects of glucocorticoids in these situations is believed to be their membrane stabilization effects that limit the development and progression of iron-dependent lipid peroxidation [15]. Recently, 21-aminosteroids or lazaroïds, a novel group of steroids that lack glucocorticoid or mineralocorticoid effects, have been shown to be potent inhibitors of iron-induced lipid peroxidation [1]. Moreover, these agents have been shown to be scavengers of lipid peroxy and phenoxy radicals [16]. In both clinical and experimental studies, these agents have been shown to reduce the severity of brain and spinal cord ischemia [9, 20].

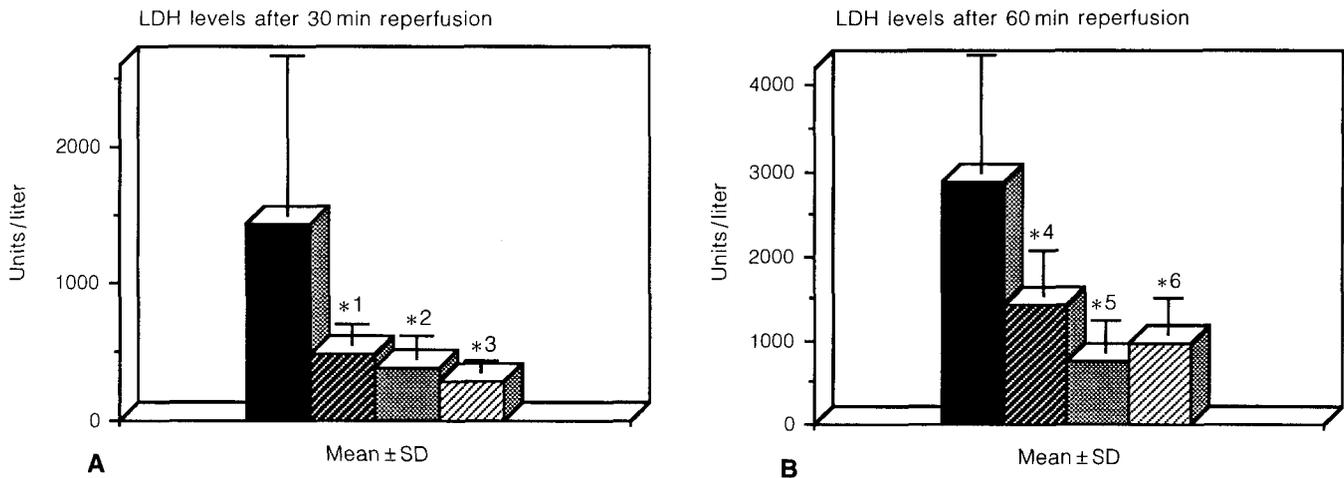


Fig. 3 A, B. The amount of LDH released into the perfusate was significantly reduced after **A** 30 and **B** 60 min, when U74500A, lidoflazine, or both drugs were added to the preservation medium. At 30 min the drug combination was more effective than U74500A alone ($P = 0.016$), whereas there was no significant difference in this comparison at 60 min. At 60 min, lidoflazine was associated with a lesser release of LDH than U74500A alone ($P = 0.037$). ■ UW solution, ▨ lidoflazine, ▩ U74500A, ▤ lidoflazine + U74500A. *1 $P = 0.01$; *2 $P = 0.009$; *3 $P = 0.002$; *4 $P = 0.016$; *5 $P = 0.004$; *6 $P = 0.003$

In the present study, the effect of adding the 21-aminosteroid U74500A to UW solution on the hepatic injury occurring as a result of cold ischemia was assessed using the isolated, perfused rat liver. The isolated, perfused liver has been shown to be a valuable tool for screening various preservation techniques, and it has recently been used extensively by us, as well as by other groups of investigators [2, 13, 14, 17–19]. In some of these studies a significant correlation between performance in the isolated, perfused liver and in vivo assessment of liver preservation has been found [12–14]. The dose of U74500A employed (30 $\mu\text{mol/l}$ or 21.6 mg/l) was within the dose range found to be effective in reducing ischemic injury to the central nervous system [9]. The magnitude of the cytoprotection achieved with U74500A was compared to that achieved with a calcium channel blocker, lidoflazine. This latter agent has previously been shown to be beneficial in this model at a dose of 5 mg/l [13], as well as in an orthotopic liver transplant model in the rat [12]. U74500A reduced the amount of hepatocellular enzymes released into the perfusate upon reperfusion after 72 h of cold storage. The improvement observed was similar to that obtained by adding lidoflazine to the UW solution.

The effect of combining a lazaroid and a calcium channel blocker was also studied. After 30 min of reperfusion, the enzyme release into the perfusate with both agents added to the UW solution was reduced, as compared to that observed when either drug was used alone ($P = 0.016$). This indicates that the two agents may have an additive effect at reducing the cell injury that occurs during ischemia, cold storage, and early reperfusion.

Since using doses greater than 5 mg of lidoflazine does not result in a greater reduction in enzyme loss with this model [13], the finding of an additional effect with U74500A and lidoflazine together suggests that the two agents work by different mechanisms to prevent cell injury. From prior studies with the lazaroids it appears as if the major action of such agents is to inhibit lipid peroxidation [4]. Another mechanism that may contribute to the cell membrane injury experienced during ischemia is the activation of phospholipases that occurs as a result of increased cytosolic Ca^{2+} levels [6]. Thus, calcium entry blockers, like lidoflazine, have been used and have been shown to reduce the cell injury associated with organ cold storage and reperfusion by limiting the entry of calcium into the cytosol.

In conclusion, in the present study the 21-aminosteroid U74500A was found to reduce the liver injury experienced during cold storage and reperfusion of rat liver in vitro. The magnitude of improvement was similar to that found with a Ca^{2+} antagonist, lidoflazine. Importantly, the effect of both agents, when used in combination, was greater than that achieved with either agent alone, at least during the early (30-min) reperfusion period.

Acknowledgements. Lidoflazine was generously donated by Kabi Pharmacia, La Jolla, California, and U74500A by Upjohn, Kalamazoo, Michigan. The authors gratefully acknowledge Dr. Rene Dumesnoy for his general support. This study was supported by grants from the Department of Veteran Affairs, the Swedish-American Foundation, and by project grant DK29961 from the National Institute of Health.

References

- Anderson DK, Saunders RD, Demediuk P, Dugan LL, Braugher JM, Hall ED, Means ED, Horrock LA (1985) Lipid hydrolysis and peroxidation in injured spinal cord: partial protection with methylprednisolone or vitamin E and selenium. *CNS Trauma* 2: 257
- Ar'Rajab A, Ahren B, Bengmark S (1991) Improved liver preservation for transplantation due to calcium channel blockade. *Transplantation* 51: 965
- Belzer FO, Southard JH (1988) Principles of solid-organ preservation by cold storage. *Transplantation* 45: 673

4. Braughler JM, Pregoner JF (1989) The 21-aminosteroid inhibitors of lipid peroxidation: reactions with lipid peroxy and phenoxy radicals. *Free Radic Biol Med* 7: 125
5. Braughler JM, Burton PS, Chase RL, Pregoner JF, Jacobsen EJ, Van Doornik FJ, Tustin JM, Ayer DE, Bundy GL (1988) Novel membrane localized iron chelators as inhibitors of iron-dependent lipid peroxidation. *Biochem Pharmacol* 37: 3853–3860
6. Chien KR, Abrams J, Serroni A, Martin JT, Farber HL (1978) Accelerated phospholipid degradation and associated membrane dysfunction in irreversible, ischemic liver cell injury. *J Biol Chem* 253: 4809
7. D'Allesandro A, Southard JH, Kalayoglu M, Belzer FO (1986) Effect of drug pretreatment on liver function following 24-hour hypothermic preservation. *Cryobiology* 23: 415
8. Hall ED (1985) High dose glucocorticoid treatment improves neurological recovery in headinjured mice. *J Neurosurg* 62: 882
9. Hall ED, Pazara KE, Braughler JM (1988) 21-aminosteroid lipid peroxidation inhibitor U74006F protects against cerebral ischemia in gerbils. *Stroke* 19: 997
10. Hall ED, Braughler JM, McCall JM (1990) Role of oxygen radicals in stroke: effect of the 21-aminosteroids (lazaroids). A novel class of antioxidants. *Prog Clin Biol Res* 361: 351–362
11. Hasselgren PO, Biber B, Fornander J (1983) Improved blood flow and protein synthesis in the postischemic liver following infusion of dopamine. *J Surg Res* 34: 44
12. Jacobsen EJ, McCall JM, Ayer DE, VanDoornik FJ, Palmer JR, Belonga KL, Braughler JM, Hall ED, Houser DJ, Krook MA (1990) Novel 21-aminosteroids that inhibit iron-dependent lipid peroxidation and protect against central nervous system trauma. *J Med Chem* 33: 1145–1151
13. Jacobsson J, Sundberg R, Valdivia LA, Starzl TE (1993) Liver preservation with lidoflazine and the University of Wisconsin solution – a dose-finding study. *Transplantation* (in press)
14. Jamieson NV, Sundberg R, Lindell S, Southard JH, Belzer FO (1988) A comparison of cold storage solutions for hepatic preservation using the isolated perfused rabbit liver. *Cryobiology* 25: 300
15. Olson LM, Klintmalm GB, Husberg BS, Nery JR, Whitten CW, Paulsen AW, McClure T (1988) Superoxide dismutase improves organ preservation in liver transplantation. *Transplant Proc* 20: 961
16. Santiago-Delpin EA, Figueroa I, Lopez R, Vasquez J (1975) Protective effects of steroids in liver ischemia. *Am Surg* 41: 683
17. Sundberg R, Lindell S, Jamieson NV, Southard JH, Belzer FO (1988) Effects of chlorpromazine and methylprednisolone on perfusion preservation of rabbit livers. *Cryobiology* 25: 417
18. Sundberg R, Ar'Rajab A, Ahren B, Bengmark S (1989) Chlorpromazine pretreatment of the donor improves liver preservation quality with UW solution in an experimental model. *Transplantation* 48: 742–744
19. Sundberg R, Ar'Rajab A, Ahren B, Bengmark S (1991) The functional effects of suppression of hypothermia-induced cell swelling in liver preservation by cold storage. *Cryobiology* 28: 150
20. Young W, Wojak JC, DeCrescito V (1988) 21-aminosteroid reduces ion shifts and edema in the rat middle cerebral artery model of regional ischemia. *Stroke* 29: 1013